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ANTINEOPLASTIC PEPTIDES

RELATED APPLICATIONS

This application is a continuation of Application No. 09/097,184, filed June 12, 1998, which is a continuation-in-part of International Application No.

- PCT/EP96/05518, filed December 11, 1996, which designated the United States, published in English, which claims priority to U.S. Patent Application No. 08/573,422, filed December 15, 1995, now abandoned. This application also claims benefit of U.S. Provisional Application entitled "Antineoplastic Peptides", which resulted from the conversion of U.S. Serial No. 08/573,422.
- The entire teachings of the above application(s) are incorporated herein by reference.

FIELD OF THE INVENTION

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The invention described herein provides novel peptides and derivatives thereof which offer potentially improved therapeutic utilities for the treatment of neoplastic diseases as compared to dolastatin -10 and -15 (U.S. Patent Nos. 4,879,276 and 4,816,444) and the compounds desribed in WO 93/23424.

SUMMARY OF THE INVENTION

Compounds of this invention include novel peptides of the formula I

R^1R^2N -CHX-CO-A-B-D-E-(G) $_s$ -K

I

	where	
5	\mathbb{R}^1	is hydrogen, methyl, or ethyl;
	\mathbb{R}^2	is methyl; or ethyl; or
	R^1 -N- R^2	together are a pyrrolidine ring;
	A	is a valyl, isoleucyl, allo-isoleucyl, 2-tert-butylglycyl, 2-ethylglycyl,
		norleucyl or norvalyl residue;
10	В	is a N-methyl-valyl, N-methyl-norvalyl, N-methyl-leucyl, N-methyl-
		isoleucyl, N-methyl-2-tert-butylglycyl, N-methyl-2-ethylglycyl, or N-
		methyl-norleucyl residue;
	D	is a prolyl, homoprolyl, hydroxyprolyl, or thiazolidine-4-carbonyl
		residue;
15	Е	is a prolyl, homoprolyl, hydroxyprolyl, thiazolidine-4-carbonyl, trans-4-
		fluoro-L-prolyl, cis-4-fluoro-L-prolyl, trans-4-chloro-L-prolyl or cis-4-
		chloro-L-prolyl residue;
	X	is ethyl, propyl, butyl, isopropyl, sec. butyl, tertbutyl, cyclopropyl, or
		cyclopentyl;
20	G	is a L-2-tert.butylglycyl, D-2-terr.butylglycyl, D-valyl, D-isoleucyl, D-
		leucyl, D-norvalyl, 1-aminopentyl-1-carbonyl, or 2,2-dimethylglycyl
		residue;
	S	is 0 or 1;
	K	is -NH- C_{1-8} -alkyl, -NH- C_{3-8} -alkenyl, -NH- C_{3-8} -alkinyl, -NH- C_{6-8} -
25		cycloalkyl, -NH- $C_{1.4}$ -alkene- $C_{3.8}$ -cycloalkyl, $C_{1.4}$ -alkyl-N- $C_{1.6}$ -alkyl, in
		which residues one CH ₂ group may be replaced by O or S, one H by
		phenyl or cyano, or 1, 2 or 3 H by F, except the N-methoxy-N-

methylamino, N-benzylamino, or N-methyl-N-benzylamino residue, or K is

$$_{10}$$
 $_{\text{NH}}$ $_{\text{NH}}$ $_{\text{NH}}$

,
$$-NH$$
 , $-NH$, $-NH$, $-NH$, $-NH$

$$-NH \xrightarrow{CH_3 CH_3} -NH \xrightarrow{N} -N \xrightarrow{E} 0$$

$$- N - NH - CH_{3} - NH - CH_{2} - CH_{2} - CH_{3}$$

$$- CH_{3} - CH_{3} - CH_{2} - CH_{2} - CH_{3}$$

and the salts thereof with physiologically tolerated acids.

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DETAILED DESCRIPTION OF THE INVENTION

In specific embodiments of the compounds of formula I, K may be -NHCH₃,

- -NHCH₂CH₃, -NH(CH₂)₂CH₃, -NH(CH₂)₃CH₃, -NH(CH₂)₄CH₃, -NH(CH₂)₅CH₃,
- -NH(CH₂)₆CH₃, -NHCH(CH₂)₇CH₃, -NHCH(CH₃)₂, -NHCH(CH₂)CH₂CH₃,
- 5 -NHCH(CH₂CH₃)₂, -NHCH(CH₂CH₂CH₃)₂, -NHC(CH₃)₃, -NHCH(CH₂CH₃)CH₂ CH₂CH₃, -NHCH(CH₃)CH(CH₃)₂, -NHCH(CH₂CH₃)CH(CH₃)₂, -NHCH(CH₃)C(CH₃)₃,
 - -NH-cyclohexyl, -NH-cycloheptyl, -NH-cyclooctyl, -N(CH₃)OCH₂CH₃,
 - -N(CH₃)OCH₂CH₂CH₃, -N(CH₃)OCH(CH₃)₂, -N(CH₃)O(CH₂)₃CH₃, -N(CH₃)OCH₂C₆H₅,
 - $-NH(CH_2)_2C_6H_5$, $-NH(CH_2)_3C_6H_5$, $-NHCH(CH_3)C_6H_5$, $-NHC(CH_3)_2C_6H_5$,
- 10 -NHC(CH₃)₂CH₂CH₃, -NHC(CH₃)(CH₂CH₃)₂, -NHCH[CH(CH₃)₂]₂, -NHC(CH₃)₂CN,
 - -NHCH(CH₂)CH(OH)C₆H₅, -NHCH₂-cyclohexyl, -NHCH₂C(CH₂)₃,
 - -NHCH₂CH(CH₃)₂, -N(CH₃)₂, -N(CH₂CH₃)₂, -N(CH₂CH₂CH₃)₂, -NHCH₂CF₃,
 - -NHCH(CH₂F)₂, -NHCH₂CH₂F, -NHCH₂CH₂OCH₃, -NHCH₂CH₂SCH₃,
 - $-NHCH_2CHCH_2$, $-NH-C(CH_3)_2CH=CH_2$, $-NHC(CH_3)_2C=CH$, $-NHC(CH_2CH_3)_2C=CH$,
- 15 -NHC(CH₃)₂CH₂CH₂OH, -NH(CH₂CH₂O)₂CH₂CH₃, -NHC(CH₃)₂CH(CH₃)₂,
 - -NHC(CH₃)₂CH₂CH₂CH₃, -NHC(CH₃)₂CH₂C₆H₅, -N(OCH₃)CH(CH₃)₂,
 - $-N(OCH_3)CH_2CH_3, -N(OCH_3)CH_2CH_2CH_3, -N(OCH_3)CH_2C_6H_5, -N(OCH_3)C_6H_5,\\$
 - $-N(CH_3)OC_6H_5$, $-NHCH[CH(CH_3)_2]_2$, $-N(OCH_3)CH_2CH_2CH_2CH_3$, or the special ring systems mentioned above.
- In one embodiment of the compounds of formula I described above, s is 0 and E is homoprolyl or hydroxyprolyl.

Preferred are compounds of the formula I where the substituents R¹, R², A, B, D, E, X, G and s have the following meanings:

- R¹ hydrogen, methyl, or ethyl, especially methyl;
- 25 R² methyl or ethyl, especially methyl;
 - A valyl, isoleucyl, 2-tert-butylglycyl, 2-ethylglycyl, norleucyl or norvalyl, especially valyl, isoleucyl, 2-tert-butylglycyl, 2-ethylglycyl,
 - B N-methyl-valyl, N-methyl-norvalyl, N-methyl-isoleucyl, N-methyl-2-tert-butylglycyl, N-methyl-2-ethylglycyl, or N-methyl-norleucyl, especially N-

methyl-valyl, N-methyl-2-ethylglycyl, N-methyl-norleucyl, N-methyl-isoleucyl, or N-methyl-2-tert.butyl-glycyl;

- D prolyl, homoprolyl or thiazolidine-4-carbonyl, especially prolyl or thiazolidine-4-carbonyl;
- prolyl, homoprolyl, thiazolidine-4-carbonyl, trans-4-fluoro-L-prolyl, cis-4-fluoro-L-prolyl, trans-4-chloro-L-prolyl or cis-4-chloro-L-prolyl, especially prolyl, trans-4-fluoro-prolyl, cis-4-fluoro-prolyl, trans-4-chloro-prolyl, or cis-4-chloro-prolyl;
- X ethyl, propyl, isopropyl, sec.butyl, tert.butyl or cyclo-propyl, especially ethyl, isopropyl, sec.butyl or tert.butyl;
 - G L-2-tert.butylglycyl, D-2-tert.butylglycyl, D-valyl, D-isoleucyl, D-leucyl or 2,2-dimethylglycyl residue;
 - s 0 or 1.

Preferred meanings for K are:

15 -NH-C₁₋₈-alkyl, -NH-C₆₋₈-cycloalkyl, -NH-CH₂-cyclohexyl, C₁₋₄-alkyl-N-C₁₋₆-alkyl, in which residues one CH₂ group may be replaced by O, one H by phenyl or 1 or 2 H by F, except the N-methoxy-N-methylamino, N-benzylamino, or N-methyl-N-benzylamino residue, or K is

$$-NH$$
 $-NH$
 $-NH$
 $-NH$

$$-NH$$
 or $-N$ NH

5 Or
$$-NH \xrightarrow{CH_3} CO - NH - CH_2 - CH_2 - CH_3$$
.

More preferred K is

 $-NHCH_{3}, -NHCH_{2}CH_{3}, -NH(CH_{2})_{2}CH_{3}, -NH(CH_{2})_{3}CH_{3}, -NH(CH_{2})_{4}CH_{3}, \\$

 $\begin{array}{ll} 10 & -\mathrm{NH}(\mathrm{CH_2})_5\mathrm{CH_3}, \ -\mathrm{NH}(\mathrm{CH_2})_6\mathrm{CH_3}, \ -\mathrm{NH}(\mathrm{CH_2})_7\mathrm{CH_3}, \ -\mathrm{NHCH}(\mathrm{CH_3})_2, \\ \\ & -\mathrm{NHCH}(\mathrm{CH_3})\mathrm{CH_2}\mathrm{CH_3}, \ -\mathrm{NHCH}(\mathrm{CH_2}\mathrm{CH_3})_2, \ -\mathrm{NHCH}(\mathrm{CH_2}\mathrm{CH_2}\mathrm{CH_3})_2, \ -\mathrm{NHC}(\mathrm{CH_3})_3, \end{array}$

 $-\mathrm{NHCH}(\mathrm{CH_2CH_3})\mathrm{CH_2CH_2CH_3}, -\mathrm{NHCH}(\mathrm{CH_3})\mathrm{CH}(\mathrm{CH_3})_2, -\mathrm{NHCH}(\mathrm{CH_2CH_3})\mathrm{CH}(\mathrm{CH_3})_2,$

 $\hbox{-NHCH}(CH_3)C(CH_3)_3, \hbox{-NH-cyclohexyl, -NH-cycloheptyl, -NH-cyclooctyl,}$

-N(CH₃)OCH₂CH₃, -N(CH₃)OCH₂CH₂CH₃, -N(CH₃)OCH(CH₃)₂,

 $15 \quad \text{-N(OCH}_3\text{)CH(CH}_3\text{)}_2, \text{-N(CH}_3\text{)OCH}_2\text{C}_6\text{H}_5, \text{-NH(CH}_2\text{)}_2\text{C}_6\text{H}_5, \text{-NH(CH}_2\text{)}_3\text{C}_6\text{H}_5,$

 $-NHCH(CH_3)C_6H_5, -NHC(CH_3)_2C_6H_5, -NHC(CH_3)_2CH_2CH_3, -NHC(CH_3)(CH_2CH_3)_2, \\$

-NHCH(CH₃)CH(OH)C₆H₅, -NHCH₂-cyclohexyl, -N(CH₃)₂, -N(CH₂CH₃)₂,

 $-N(CH_2CH_3CH_3)_2$, $-NHCH(CH_2F)_2$, $-NHC(CH_3)CH = CH_2$, $-NHC(CH_3)_2CN$,

 $-{\rm NHC}({\rm CH_3})_2{\rm C} \equiv {\rm CH, -NHC}({\rm CH_3})_2{\rm CONH_2, -NHCH}[{\rm CH}({\rm CH_3})_2]_2, -{\rm N}({\rm OCH_3}){\rm CH_2C_6H_5},$

 $20 \quad \text{-N(OCH}_3\text{)CH}_2\text{CH}_3, \text{-N(OCH}_3\text{)CH}_2\text{CH}_2\text{CH}_3, \text{-N(OCH}_3\text{)CH}_2\text{CH}_2\text{CH}_2\text{CH}_3,$

In one embodiment of the preferred compounds of formula I described above, s is 0 and E is homoprolyl or hydroxyprolyl.

Especially preferred are compounds of the formula I where

 R^1 and R^2 are methyl,

5 A is a valyl, isoleucyl, 2-tert.-butylglycyl residue

B is a N-methylvalyl, N-methyl-isoleucyl, N-methyl-2-tert.-butylglycyl residue,

D is a prolyl or thiazolidine-4-carbonyl residue,

E is a prolyl, cis-4-fluoro-L-prolyl, or cis-4-chloro-L-prolyl residue,

X is a isopropyl, sec.-butyl, or tert.-butyl residue,

10 s is 0, and

K is

 $\hbox{-NHCH}(CH_3)_2, \hbox{-NHCH}(CH_3)CH_2CH_3, \hbox{-NHCH}(CH_2CH_3)_2, \hbox{-NHCH}(CH_2CH_2CH_3)_2,$

-NHC(CH₃)₃, -NHCH(CH₂CH₃)CH₂CH₂CH₃, -NHCH(CH₃)CH(CH₃)₂,

 $-\mathrm{NHCH}(\mathrm{CH_2CH_3})\mathrm{CH}(\mathrm{CH_3})_2, \ \ -\mathrm{NHCH}(\mathrm{CH_3})\mathrm{C}(\mathrm{CH_3})_3, \ -\mathrm{NH-cycloheptyl}, \ -\mathrm{NH-cyclooctyl},$

15 -N(CH₃)OCH₂CH₃, -N(CH₃)OCH₂CH₂CH₃, -N(CH₃)OCH(CH₃)₂, -N(OCH₃)CH(CH₃)₂,

 $-N(CH_3)OCH_2C_6H_5$, $-NH(CH_2)_2C_6H_5$, $-NH(CH_2)_3C_6H_5$, $-NHCH(CH_3)C_6H_5$,

 $-NHC(CH_{3})_{2}C_{6}H_{5},-NHC(CH_{3})_{2}CH_{2}CH_{3},-NHC(CH_{3})(CH_{2}CH_{3})_{2},\\$

 $\hbox{-NHCH}(CH_3)CH(OH)C_6H_5, \hbox{-NHCH}(CH_2F)_2, \hbox{-NHC}(CH_3)_2CH_2CH_2OH,$

 $-{\rm NH}({\rm CH_2CH_2O})_2{\rm CH_2CH_3}, -{\rm NHC}({\rm CH_3})_2{\rm CH} = {\rm CH_2}, \ -{\rm NHC}({\rm CH_3})_2{\rm CH}({\rm CH_3})_2,$

 $20 \quad \text{-N(OCH}_3\text{)CH}_2\text{CH}_3, \text{-N(OCH}_3\text{)CH}_2\text{CH}_2\text{CH}_3, \text{-N(OCH}_3\text{)CH}_2\text{CH}_2\text{CH}_2\text{CH}_3,$

 $-\mathrm{NHC}(\mathrm{Ch}_3)_2\mathrm{CN}, -\mathrm{NHC}(\mathrm{CH}_3)_2\mathrm{C} = \mathrm{CH}, -\mathrm{NHCH}[\mathrm{CH}(\mathrm{CH}_3)_2]_2, -\mathrm{NHC}(\mathrm{CH}_3)_2\mathrm{CONH}_2,$

-NHC(CH₃)₂CH₂C₆H₅, -N(OCH₃)C₆H₅, -N(OCH₃)CH₂C₆H₅,

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$$-NH$$
 $-NH$
 $-NH$

This invention also provides methods for preparing the compounds of formula I, pharmaceutical compositions containing such compounds together with a pharmaceutically acceptable carrier and methods for using same for treating cancer in mammals.

The new compounds may be present as salts with physiologically tolerated acids such as: hydrochloric acid, citric acid, tartaric acid, lactic acid, phosphoric acid, methanesulfonic acid, acetic acid, formic acid, maleic acid, fumaric acid, malic acid, succinic acid, malonic acid, sulfuric acid, L-glutamic acid, L-aspartic acid, pyruvic acid, mucic acid, benzoic acid, glucuronic acid, oxalic acid, ascorbic acid and acetylglycine.

Thus, the peptides can be assembled sequentially from amino acids or by linking suitable small peptide fragments. In the sequential assemblage, starting at the C terminus the peptide chain is extended stepwise by one amino acid each time. In fragment coupling it is possible to link together fragments of different lengths, and the fragments in turn can be obtained by sequential assemblage from amino acids or themselves by fragment-coupling.

Both in the sequential assemblage and in the fragment coupling it is necessary to link the units by forming an amide linkage. Enzymatic and chemical methods are suitable for this.

Chemical methods for forming the amide linkage are described in detail by Mueller, Methoden der organischen Chemie Vol. XV/2, pp 1 to 264, Thieme Verlag, Stuttgart, 1974; Stewart, Young, Solid Phase Peptide Synthesis, pp 31 to 34, 71 to 82,

2-hydroxypyridine.

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Pierce Chemical Company, Rockford, 1984; Bodanszky, Klausner, Ondetti, Peptide Synthesis, pp 85 to 128, John Wiley & Sons, New York, 1976; The Practice of Peptide Synthesis, M. Bodanszky, A. Bodanszky, Springer-Verlag, 1994, and other standard works on peptide chemistry. Particular preference is given to the azide method, the symmetric and mixed anhydride method, in situ generated or preformed active esters, the use of urethane protected N-carboxy anhydrides of amino acids and the formation of the amide linkage using coupling reagents, especially dicyclohexylcarbodiimide (DCC), diisopropylcarbodiimide (DIC), 1-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ), pivaloylchloride, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDCI), n-propanephosphonic anhydride (PPA), N,N-bis(2-oxo-3-10 oxazolodinyl)-amidophosphoryl chloride (BOP-C1), bromo-tris-pyrrolidinophosphonium hexafluorophosphate (PyBrop), diphenylphosphoryl azide (DPPA), Castro's reagent (BOP, PyBop), O-benzotriazolyl-N,N,N',N'-tetramethyluronium salts (HBTU), O-azabenzotriazolyl-N,N,N',N'-tetramethyluronium salts (HATU), diethylphosphoryl cyanide (DEPCN), 2,5-diphenyl-2,3-dihydro-3-oxo-4hydroxythiophene dioxide (Steglich's reagent; HOTDO) and 1,1'-carbonyldiimidazole (CDI). The coupling reagents can be employed alone or in combination with additives such as N.N-dimethyl-4-aminopyridine (DMAP), N-hydroxy-benzotriazole (HOBt), N-

Whereas it is normally possible to dispense with protective groups in enzymatic peptide synthesis, reversible protection of reactive groups not involved in formation of the amide linkage is necessary for both reactants in chemical synthesis. Three conventional protective group techniques are preferred for the chemical peptide synthesis: the benzyloxycarbonyl (Z), the t-butoxycarbonyl (Boc) and the 9-fluorenylmethoxycarbonyl (Fmoc) techniques.

hydroxybenzotriazine (HOOBt), Azabenzotriazole, N-hydroxysuccinimide (HOSu) or

Identified in each case is the protective group on the alpha-amino group of the chain-extending unit. A detailed review of amino-acid protective groups is given by Mueller, Methoden der organischem Chemie vol. XV/1, pp 20 to 906, Thieme Verlag,

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Stuttgart, 1974. The units employed for assembling the peptide chain can be reacted in solution, in suspension or by a method similar to that described by Merrifield in J. Amer. Chem. Soc. 85 (1963) 2149.

Suitable for peptide synthesis in solution are all solvents which are inert under the reaction conditions, especially water, N,N-dimethylformamide (DMF), dimethyl sulfoxide (DMSO), acetonitrile, dichloromethane (DCM), ethyl acetate, 1,4-dioxane, tetrahydrofuran (THF), N-methyl-2-pyrrolidone (NMP) and mixtures of the said solvents.

Peptide synthesis on the polymeric support can be carried out in all inert organic solvents in which the amino-acid derivatives used are soluble. However, preferred solvents additionally have resin-swelling properties, such as DMF, DCM, NMP, acetonitrile and DMSO, and mixtures of these solvents. After synthesis is complete, the peptide is cleaved off the polymeric support. The conditions under which cleavage off the various resin types is possible are disclosed in the literature. The cleavage reactions most commonly used are acid- and palladium-catalyzed, especially cleavage in liquid anhydrous hydrogen fluoride, in anhydrous trifluoromethanesulfonic acid, in dilute or concentrated trifluoroacetic acid, palladium-catalyzed cleavage in THF or THF-DCM mixtures in the presence of a weak base such as morpholine or cleavage in acetic acid/dichloromethane/ trifluoroethanol mixtures. Depending on the chosen protective groups, these may be retained or likewise cleaved off under the cleavage conditions.

Partial deprotection of the peptide may also be worthwhile when certain derivatization reactions are to be carried out.

Peptides dialkylated at the N-terminus can be prepared either by coupling on the appropriate N,N-di-alkylamino acids in solution or on the polymeric support, by reductive alkylation of the resin-bound peptide in DMF/1% acetic acid with NaCNBH₃ and the appropriate aldehydes, by hydrogenation of the peptide in solution in the presence of aldehyde or ketone and Pd/C.

The various non-naturally occurring amino acids as well as the various nonamino acid moieties disclosed herein may be obtained from commercial sources or

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synthesized from commercially-available materials using methods known in the art. For example, amino acids building blocks with R¹ and R² moieties can be prepared according to E. Wuensch, Houben Weyl, Meth. d. Org. Chemie, Bd. XV, 1, p. 306 following, Thieme Verlag Stuttgart 1974 and Literature cited therein.

The compounds of this invention may be used to inhibit or otherwise treat solid tumors (e.g. tumors of the lung, breast, colon, prostate, bladder, rectum, or endometrial tumors) or hematological malignancies (e.g. leukemias, lymphomas) by administration of the compound to the mammal.

It is a special advantage of the new compounds that they are very resistant to enzymatic degradation and can also be administered orally.

Administration may be by any of the means which are conventional for pharmaceutical, preferably oncological, agents, including oral and parenteral means such as subcutaneously, intravenously, intramuscularly and intraperitoneally.

The compounds may be administered alone or in the form of pharmaceutical compositions containing a compound of formula I together with a pharmaceutically accepted carrier appropriate for the desired route of administration. Such pharmaceutical compositions may be combination products, i.e., may also contain other therapeutically active ingredients.

The dosage to be administered to the mammal with contain an effective tumor-inhibiting amount of active ingredient which will depend upon conventional factors including the biological activity of the particular compound employed; the means of administration; the age, health and body weight of the recipient; the nature and extent of the symptoms; the frequency of treatment; the administration of other therapies; and the effect desired. A typical daily dose will be about 0.05 to 50 milligrams per kilogram of body weight on oral administration and about 0.01 to 20 milligrams per kilogram of body weight on parenteral administration.

The novel compounds can be administered in conventional solid or liquid pharmaceutical administration forms, e.g. uncoated or (film-)coated tablets, capsules, powders, granules, suppositories or solutions. These are produced in a conventional

manner. The active substances can for this purpose be processed with conventional pharmaceutical aids such as tablet binders, fillers preservatives, tablet disintegrants, flow regulators, plasticizers, wetting agents, dispersants, emulsifiers, solvents, sustained release compositions, antioxidants and/or propellant gases (cf. H. Sucker et al.:

5 Pharmazeutische Technologie, Thieme-Verlag, Stuttgart, 1978). The administration forms obtained in this way normally contain 1-90% by weight of the active substance.

The following examples are intended to illustrate the invention. The proteinogenous amino acids are abbreviated in the examples using the known three-letter code. Other abbreviations used: $Me_2Val = N,N-dimethylvaline, MeVal = N$

10 N-methylvaline.

EXAMPLES

A. General procedures

I. The peptides claimed in claim 1 are either synthesized by classical solution synthesis using standard Z- and Boc-methodology as described above or by standard methods of solid-phase synthesis using Boc and Fmoc protective group techniques.

In the case of solid phase synthesis, the N,N-dialkylpenta- or hexapeptide acids are liberated from the solid support and further coupled with the corresponding C-terminal amines in solution. BOP-C1 and PyBrop were used as reagents for coupling of the amino acid following the N-methylamino acids. The reaction times were correspondingly increased. For reductive alkylation of the N-terminus, the peptide-resin was deprotected at the N terminus and then reacted with a 3-fold molar excess of aldehyde or ketone in DMF/1% acetic acid with addition of 3 equivalents of NaCNBH₃. After the reaction was complete (negative Kaisertest) the resin was washed several times with water, isopropanol, DMF and dichloromethane.

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In solution synthesis, the use of either Boc-protected amino acid NCAs (N-tert.-butyloxycarbonyl-amino acid-N-carboxy-anhydrides), Z-protected amino acid NCAs (N-benzyloxycarbonyl-amino acid-N-carboxy-anhydrides), or the use of pivaloylchloride as condensing agent respectively is most advantageous for coupling of the amino acid following the N-methylamino acids. Reductive alkylation of the N terminus can e.g. be achieved by reaction of the N-terminally deprotected peptides or amino acids with the corresponding aldehydes or ketones using NaCNBH₃ or hydrogen, Pd/C.

Purification and characterization of the peptides

Purification was carried out by gel chromatography (SEPHADEX G-10, G15/10% HOAc, SEPHADEX LH20/MeOH), medium pressure chromatography
(stationary phase: HD-SIL C-18, 20-45 mikron, 100 Angstrom; mobile phase:
gradient with A = 0.1% TFA/MeOH, B = 0.1% TFA/water), or preparative

HPLC (stationary phase: Waters Delta-Pak C-18, 15 mikron, 100 Angstrom;
mobile phase: gradient with A = 0.1% TFA/MeOH, 3 = 0.1% TFA/water).

The purity of the resulting products was determined by analytical HPLC (stationary phase: 100 2.1 mm VYDAC C-18, 5 1, 300 A; mobile phase: acetonitrile-water gradient, buffered with 0.1% TFA, 40°C).

Characterization was by amino-acid analysis and fast atom bombardment mass spectroscopy.

B. Specific procedures

EXAMPLE 1 (SEQ ID NO: 1)

Me₂Val-Val-MeVal-Pro-Pro-NHCH(CH₃)₂

- a) Z-MeVal-Pro-OME
- 5 66.25 g (250 mmol) Z-MeVal-OH were dissolved in 250 ml dry dichloromethane. After addition of 36.41 ml (262.5 mmol) triethylamine, the reaction mixture was cooled to -25° C and 32.27 ml (262.5 mmol) pivaloyl chloride were added. After stirring for 2,5 h, 41.89 g (250 mmol) H-Pro-OMe x Chl in 250 ml dichloromethane, neutralized with 36.41 ml (262.5 mmol) triethylamine at 0°C, were added to the reaction mixture. Stirring continued for 2 h at -25° C and overnight at room temperature. The reaction mixture was diluted with dichloromethane and thoroughly washed with saturated aqueous NaHCO₃ solution (3x), water (1x), 5 % citric acid (3x) and saturated NaCl solution. The organic phase was dried over sodium sulfate and evaporated to dryness. The residue (91.24 g) was stirred with petroleum ether overnight and filtered. 62.3 g of product were obtained.
- b) H-MeVal-Pro-OMe
 48.9 g (130 mmol) Z-MeVal-Pro-OMe were dissolved in 490 ml methanol.
 20 After addition of 10.9 ml (130 mmol) concentrated hydrochloric acid and 2.32 g
 10 % Palladium/charcoal, the reaction mixture was hydrogenated. Filtration and evaporation to dryness yielded 36.32 g of the product.
 - c) Z-Val-MeVal-Pro-OMe
- 18.1 g (65 mmol) H-MeVal-Pro-OMe, 21.6 g (78 mmol) Z-Val-N-carboxyanhydride and 22.8 ml (130 mmol) diisopropylethylamine were stirred in 110 ml DMF at 40° C for 2 d. After evaporation of DMF, dichloromethane was added and the organic phase washed with saturated aqueous NaHCO₃ solution (3x), water (1x), 4 % citric acid (3X) and saturated NaCl solution. The organic

phase was dried over sodium sulfate and evaporated to dryness. The product (29.3 g) was obtained as a viscous oil.

- d) H-Val-MeVal-Pro-OMe
- 29.3 g (61.6 mmol) of Z-Val-MeVal-Pro-OMe were dissolved in 230 ml methanol. After addition of 1.15 g 10% Palladium/charcoal, the reaction mixture was hydrogenated. Filtration and evaporation to dryness yielded 21.96 g of the product.
- 2-Val-Val-MeVal-Pro-OMe
 15.29 g (61 mmol) Z-Val-OH and 21.96 g (61 mmol) H-Val-MeVal-Pro-OMe were dissolved in 610 ml dichloromethane and cooled to 0°C. After addition of 8.16 ml (73.2 mmol) N-Methylmorpholine, 2.77 g (20.3 mmol) HOBt and 11.73 g (61 mmol) EDCI, the reaction mixture was stirred overnight at room temperature, diluted with dichloromethane and thoroughly washed with saturated aqueous NaHCO₃ solution (3x), water (1x), 5 % citric acid (3X) and saturated NaCl solution. The organic phase was dried over sodium sulfate and evaporated to dryness to yield 31.96 g of the product.
- 20 f) Z-Val-Val-MeVal-Pro-OH

 31.96 g (57 mmol) Z-Val-Val-MeVal-Pro-OMe were dissolved in 250 ml
 methanol. 102.6 ml of a 1 N LiOH solution was added and the mixture stirred
 overnight at room temperature. After addition of 500 ml water, the aqueous
 phase was washed three times with ethyl acetate, adjusted to pH 2 at 0° C and
 extracted three times with ethyl acetate. The organic phase was dried over
 sodium sulfate and evaporated to dryness yielding 30.62 g of the desired product
 as a white solid.

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- Z-Val-Val-MeVal-Pro-Pro-NHCH(CH₃)₂
 2 g (3.35 mmol) Z-Val-Val-MeVal-Pro-OH and 0.664 g (3.35 mmol) H-Pro-NHCH(CH₃)₂ were dissolved in 34 ml of dry dichloromethane. After cooling to 0° C, 1.35 ml (12.1 mmol) N-methylmorpholine, 0.114 g (0.84 mmol) HOBt and 0.645 g (3.35 mmol) EDCI were added and the reaction mixture stirred overnight at room temperature. 80 ml dichloromethane were added and the organic phase thoroughly washed with saturated aqueous NaHCO₃ solution (3x), water (1x), 5 % citric acid (3x) and saturated NaCl solution (1x). The organic phase was dried over sodium sulfate and evaporated to dryness to yield 1.96 g of the product which was used in the next reaction without further purification.
 - h) Me₂Val-Val-MeVal-Pro-Pro-NHCH(CH₃), 1.96 g Z-Val-Val-MeVal-Pro-Pro-NHCH(CH₃), were dissolved in 11 ml methanol. 0.054 g 10 % Pd/C were added under nitrogen atmosphere and the reaction mixture hydrogenated at room temperature for 4 h. After addition of $0.86~\mathrm{ml}$ (11.24 mmol) of a 37 % aqueous formal dehyde solution and 0.281 g 10% Pd/C, hydrogenation was continued for 5 h. Filtration and evaporation of the solvent gave rise to 2.77 g of crude product. Further purification was achieved by dissolving the peptide in water, adjusting the pH to 2 and extracting the aqueous phase three times with ethyl acetate. The aqueous phase was then adjusted to pH 8-9 and extrcted four times with dichloromethane. The organic phase was dried over sodium sulfate to yhield 1.37 g of purified product as a white foam. The compound was further purified using medium pressure liquid chromatography (10 - 50 % A in 10 min.; 50 - 90 % A in 320 min.). Fractions containing the product were combined, lyophilized, redissolved in water and the pH adjusted to 9 with 1 N LiOH. After extraction with dichloromethane, the organic phase was dried over sodium sulfate and evaporated to dryness. Lyophilization led to 500 mg of pure product, which was characterized by fast atom bombardment mass spectrometry ($[M+H]^+ = 593$).

EXAMPLE 2 (SEQ ID NO: 1)

Me₂Val-Val-MeVal-Pro-Pro-NHC(CH₃)₃

- Z-Val-Val-MeVal-Pro-Pro-NHC(CH₃)₃
 2 g (3.35 mmol) Z-Val-Val-MeVal-Pro-OH and 0.692 g (3.35 mmol) H-Pro-NHC (CH₃)₃ were dissolved in 34 ml of dry dichloromethane. After cooling to 0°C, 1.35 ml (12.1 mmol) N-methylmorpholine, 0.114 g (0.84 mmol) HOBt and 0.645 g (3.35 mmol) EDCI were added and the reaction mixture stirred overnight at room temperature. 80 ml dichloromethane were added and the organic phase thoroughly washed with saturated aqueous NaHCO₃ solution (3x), water (1x), 5
 % citric acid (3x) and saturated NaCl solution (1x). The organic phase was dried over sodium sulfate and evaporated to dryness to yield 1.8 g of the product which was used in the next reaction without further purification.
- k) Me₂Val-Val-MeVal-Pro-Pro-NHC(CH₂)₃ 15 1.8 g Z-Val-Val-MeVal-Pro-Pro-NHC(CH₃)₃ were dissolved in 10 ml methanol. 0.049 g 10 % Pd/C were added under nitrogen atmosphere and the reaction mixture hydrogenated at room temperature for 4 h. After addition of 0.86 ml (11.24 mmol) of a 37 % aqueous formaldehyde solution and 0.252 g 10 % Pd/C, hydrogenation was continued for 5 h. Filtration and evaporation of the solvent 20 gave rise to 1.82 g of crude product. The compound was further purified using medium pressure liquid chromatography 910 - 50 % A in 10 min.; 50 - 90 % A in 320 min.). Fractions containing the product were combined, lyophilized, redissolved in water and the pH adjusted to 9 with 1 N LiOH. After extraction with dichloromethane, the organic phase was dried over sodium sulfate and 25 evaporated to dryness. Lyophilization led to 547 mg of pure product, which was characterized by fast atom bombardment mass spectrometry ($[M+H]^+ = 607$).

The following compounds were prepared or can be prepared according to examples 1 and 2:

	3.	Xaa	. Val	. Xab	Pro	Xac
5	4.	Xaa	. Val	. Xab	Pro	Xad
	5.	Xaa	Val	. Xab	Pro	Xae
	6.	Xaa	Val	Xab	Pro	Xaf
	7.	Xaa	Val	Xab	Pro	Xag
	8.	Xaa	Val	dsX	Pro	Xah
	9.	Xaa	Val	Xab	Pro	Xai
	10.	Xaa	Val	dsX	Pro	Xak
10	11.	Xaa	Val	Xab	Pro	Xal
~ ~	12.	Xaa	Val	Xab	Pro	Xam
	13.	Xaa	Val	Xab	Pro	Xan
	14.	Xaa	Val	Xab	Pro	Xao
	15.	Xaa	Val	daX	Pro	Xap
	15.	Xaa	Val	طهX	Pro	Xaq
	17.	Xaa	Val	daX	Pro	Xar
15	18.	Xaa	Val	ظة٪	Pro	Xas
13	19.	Xaa	Val	dsX	Pro	Xat
	20.	Xaa	Val	ظهلا	Pro	Xau
	21.	Xaa	Val	dsX	Pro	Xav
	22.	Xaa	Val	dsX	Pro	Xaw
	23.	Xaa	Val	daX	Pro	Xax
	24.	Xđđ	Val	dsX	Pro	Xay
20	25.	Xaa	Val	dsX	Pro	Xaz
20	26.	Xaa	Val	Xab	Pro	adX
	27.	Xaa	Val	Kab	Pro	ddX
	28.	Xaa	Val	Dax	Pro	Xay
	29.	Xaa	Val	Xab	Pro	XbdX
	30.	Xaa	Val	Xab	Pro	Xbe
	31.	Xaa	Val	Xab	Pro	Xbf
25	32.	Xaa	Val	Xab	Pro	Xbg
23	33.	Xaa	Val	Xab	Pro	Xbh
	34.	Xaa	Val	Xab	Pro	Xbi
	35.	Xaa	Val	Xab	Pro	Xbk
	36.	Xaa	Val	Xab	Pro	Xbl
	37.	Xaa	Val.	Xab	Pro	Xbm
	38.	Xaa	Val	Xab	Pro	Xbn

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39. Xaa Val Xab Pro Xbo
         40. Xaa Val Xab Pro Xbp
         41. Xaa Val Xab Pro Xbq
         42. Xaa Val Xab Pro Xbr
        43. Xaa Val Xab Pro Xbs
        44. Xaa Val Xab Pro Xbt
5
        45. Xaa Val Xab Pro Xbu
        46. Xaa Val Xab Pro Xbv
        47. Xaa Val Xab Pro Xbw
        48. Xaa Val Xab Pro Xbx
        49. Xaa Val Xab Pro Xbv
        50. Xaa Val Xab Pro Xbz
        51. Xaa Val Xab Pro Xca
10
        52. Xaa Val Xab Pro Xcb
        53. Xaa Val Xab Pro Xcc
        54. Xaa Val Xab Pro Xcd
        55. Xaa Val Xab Pro Xce
        56. Xaa Val Xab Pro Xcf
        57. Xaa Xdf Xab Pro Xay
        58. Xaa Val Xab Pro Xch
15
        59. Xaa Val Xab Pro Xci
        60. Xaa Val Xab Pro Xck
        61. Xaa Val Xab Pro Xcl
        62. Xaa Val Xab Pro Xcm
        63. Xaa Val Xab Pro Xon
        64. Xaa Val Xab Pro Xco
        65. Xaa Val Xab Pro Xcp
20
        66. Xaa Val Xab Pro Xcq
        67. Xaa Val Xab Pro Xcr
        68. Xaa Val Xab Pro Xcs
        69. Xaa Val Xab Pro Xct
        70. Xaa Val Xab Pro Xcu
       71. Xcw Val Xab Pro Xcv
       72. Xcx Val Xab Pro Xcv
25
       73. Xaa Val Xab Pro Pro Xcy
       74. Xaa Val Xab Pro Pro Xcz
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75. Xaa Val Xda Pro Xcv 76. Xaa Xdb Xab Pro Xcv 77. Xdc Val Xab Pro Xcv 78. Xaa Ile Xab Pro Xcv 79. Xdd Val Xab Pro Xcv 80. Xde Val Xab Pro Xcv 5 81. Xaa Xdf Xab Pro Xcv 82. Xaa Val Xab Pro Xcg 83. Xaa Val Xab Pro Pro Xdg 84. Xaa Val Xab Pro Pro Xdh 85. Xaa Val Xab Pro Pro Xdi 86. Xaa Val Xab Pro Pro Xdk 87. Xaa Val Xdl Pro Xcv 10 88. Xde Val Xab Pro Xay 89. Xaa Val Xdl Pro Xay 90. Xaa Val Xab Pro Xdm 91. Xaa Val Xab Pro Xdn 92. Xaa Val Xab Pro Xdo 93. Xaa Val Xab Pro Xdp 94. Xaa Val Xab Pro Xdq 15 95. Xaa Val Xab Pro Pro Xdr 96. Xaa Val Xab Pro Xds 97. Xaa Val Xbc Pro Xcv 98. Xaa Ile Xab Pro Xay 99. Xcw Val Xab Pro Xay 100. Xaa Val Xbc Pro Xal 101. Xaa Val Xdl Pro Xal 20 102. Xaa Xdf Xab Pro Xal 103. Xaa Ile Xab Pro Xal 104. Xdd Val Xab Pro Xal 105. Xde Val Xab Pro Xal 106. Xcx Val Xab Pro Xcy 107. Xcw Val Xab Pro Xal 108. Xcx Val Xab Pro Xal 25 109. Xcw Val Xab Pro Xav 110. Xcx Val Xab Pro Xav 111. Xcw Val Xab Pro Xaw 112. Xcx Val Xab Pro Xaw 113. Xab Val Xab Pro Xay

114. Xab Val Xab Pro Xcv

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115. Xab Val Xab Pro Xal
         116. Xab Val Xab Pro Xam
         117. Xab Val Xab Pro Xan
         118. Xab Val Xab Pro Xao
         119. Xab Val Xab Pro Xav
         120. Xab Val Xab Pro Xaw
 5
         121. Xab Val Xab Pro Xat
         122. Xab Val Xab Pro Xau
        123. Kab Val Kab Pro Kbf
        124. Xab Val Xab Pro Xbm
        125. Xab Val Xab Pro Xbn
        126. Xab Val Xab Pro Xbo
        127. Xab Val Xab Pro Xch
10
        128. Xaa Val Xab Pro Xdt
        129. Xaa Val Xab Pro Xdu
        130. Xaa Val Xab Pro Xdv
        131. Xaa Val Xab Pro Xdw
        132. Xaa Val Xab Pro Xdx
        133. Xaa Val Xab Pro Xdy
        134. Xaa Val Xab Pro Xdz
15
        135. Xaa Val Xab Pro Xea
       136. Xaa Val Xab Pro Xeb
       137. Xaa Val Xab Pro Xec
       138. Xaa Val Xab Pro Xed
       139. Xaa Val Xab Pro Xef
       140. Xaa Val Xab Pro Xeg
       141. Xaa Val Xab Pro Xeh
20
       142. Xaa Val Xab Pro Xei
       143. Xaa Val Xab Pro Xek
       144. Xaa Val Xab Pro Xel
       145. Xaa Val Xab Pro Xem
       146. Xaa Val Xab Pro Xen
       147. Xaa Val Xab Pro Xeo
       148. Xaa Val Xab Pro Xep
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       149. Xaa Val Xab Pro Xeq
      150. Xaa Val Xab Pro Xer
      151. Xaa Val Xab Pro Xcg
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Examples for the MS-characterization of the synthesized novel compounds are given in the following table.

5	EXAMPLE	Fast atom bombardment MS analy-
3	[No.]	[MolWeight (measured)]
	3.	563
	4.	579
	5.	593
	6.	607
10	7.	621
10	8.	635
	11.	607
	12.	607
	13.	621
	14.	649
15	15.	635
13	16.	635
	17.	635
	18.	635
	19.	621
	20.	621
20	21.	635
20	22.	635
	25.	633
	26.	647
	27.	661
	31.	623
25	32.	671
20	33.	667
	34.	531
	35.	655
	36.	655
	37.	669

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	3	ä
	7	::
	7	:
	8 8	thur):
	33 50 51	11. divide
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		there there are that

	EXAMPLE	Fast atom bombardment MS analy sis.	r -
	38.	621	
	39.	635	
	41.	649	
5	42.	621	
	43.	633	
	44.	667	
	45.	607	
	46.	647	
10	47.	668	
10	48.	655	
	49.	669	
	50.	685	
	51.	629	
	52.	625	
1.5	53.	721	
15	5 5.	579	
	58.	623	
	61.	597	
	62.	621	
	63.	609	
20	64.	625	
20	65.	635	
	66.	591	
	67.	715	
	68.	685	
	69.	685	
25	70.	591	
23	71.	607	
	72.	621	
	74.	706	
	75.	579	
	76.	579	
	7 7.	579	

	EXAMPLE	Past ator	n bombar	dment	MS	analy-	
	73.	507					
	79.	607					
	80.	607					
5	81.	607					
	82.	637					
	83.	692					
	84.	706					
	85.	706					
	86.	706					
10	87.	607					
	90.	635					
	92.	659					
	93.	617					
	94.	636					
	95.	678					
15	128.	671					
	131.	625					
	139	625					
	151.	637					
20	Table I - Sequence Ide to Examples 1 and 2 Compound Number(s)	entificati	on of Co Sequen				
	Compound Number (2)		pedaen	ice ib	TACT!	mer	
	1-56, 58-72, 75, 77, 79, 80, 82, 1 87-94, 96, 97, 99-101, 104-151						
	73, 74, 83-86, 95,			2			
	57, 76, 81, 102			3			
25	78, 98, 103			4			

The symbols Xaa in the summary have the following meanings:

Xaa: N,N-Dimethylvaline
Xab: N-Methylvaline

Xac:

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Xad:

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Xae:

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Xaf:

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Xag:

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N NH CH₃

N—NH CH₃

NH CH

NH CH₃

NH CH3

The Residence of the Royal Street, and also the Street of Street o

Xah:

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Xai:

10 Xak:

15 Xal:

Xam: 20

Xan:

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Xao:

5

Xap:

10

Xaq:

15

Xar:

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Xas:

Xat:

5

Xau:

10

Xav:

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Xaw:

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Xax:

CH₃
CH₃
CH₃

Xay:

5

Xaz:

10

15

Xbb:

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Xbc:

N-Methyl-isoleucine

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Xbe:

5

Xbf:

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Xbg:

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If it is fiften to be it is in a constraint to the form of fiften is fiften in the first to the

: ndX

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xbi:

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Xbk:

Xbl:

O H3C

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:mdX

NH CH3

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Xbn:

NH CH3 CH3

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The state of the s

Xbq: 5 Xbr: 10 Xbs: 15 Xbt: Xbu: 20

Xbv:

Xbw

Xcb: CH₃ C 5 Proline adamantyl(1)amide Xcc: Xcd: 10 ·CH3 Xce: 15 Xcf: 20 Xcg:

-36-

Xcm:

Xco:

5

10 xcp:

15 Xcq:

20 Xcr:

25

Xcs:

Xcu:

Xc::

10

5

Xcv:

15

Xcw:

N-Methyl-N-ethyl-valine

Xcx:

N, N-Diethylvaline

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Xcy:

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XCI:

H. Hard Tries for H. H. H. Lenn S. S. C. House H. House H. House H. H. Harre J. Harre J

Xda: N-Metny1-2-aminobutyroyl

Xdb: 2-aminobutyroyl

Xdc: N, N-Dimethyl-2-aminobutyroyl

5 Xdd: N,N-Dimethyl-2-tert.butylglycine

Xde: N, N-Dimethyl-isoleucine

Xdf: 2-tert.butylglycine

15
$$xdh:$$
 $H_3C \longrightarrow CH_3$ $NH \longrightarrow CH_3$ CH_3

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$$\text{Xdi:} \\ \text{H}_3\text{C} \\ \text{NH} \\ \text{CH}_3 \\ \text{O} \\ \text{CH}_3$$

Xdm:

5

Xdn:

$$\begin{array}{c|c} & & & \\ & & & \\ NH & & C = N \\ \hline \\ CH_3 & & \\ \end{array}$$

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Xdo:

15

Xdp:

20

Xdq:

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Xdr:

Xds:

...

Xdt:

5 Xdu:

Xdv:

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Xdw:

15

Xdx:

20

Xdy:

25

Xdz:

-40-

CH₃

Xeg:

5

Xeh:

10 **Xei**:

Xek:

15

20 Xel:

Xem:

25

Xan:

20

Xeo:

$$\text{Xep:} \qquad \qquad \begin{array}{c} \text{CH}_3 \\ \text{CH}_3 \\ \end{array}$$

$$C1$$
 CH_3
 NH
 CH_3
 CH_3
 CH_3

Compounds of this invention may be assayed for anti-cancer activity by conventional methods, including for example, the methods described below.

A. In vitro methodology

Cytotoxicity was measured using a standard methodology for adherent cell lines such as the microculture tetrazolium assay (MTT). Details of this assay have been published (Alley, MC et al, Cancer Research 48:589-601, 1988).

Exponentially growing cultures of tumor cells such as the HT-29 colon carcinoma or LX-1 lung tumor are used to make microtiter plate cultures. Cells are seeded at 3000 cells per well in 96-well plates (in 150 µl or media), and grown overnight at 37°C. Test compounds are added, in 10-fold dilutions

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varying from 10^{-4} M to 10^{-10} M. Cells are then incubated for 72 hours. To determine the number of viable cells in each well, the MTT dye is added (50 μ l or 3 mg/ml solution of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide in saline). This mixture is incubated at 37°C for 5 hours, and then 50 μ l of 25% SDS, pH2 is added to each well. After an overnight incubation, the absorbance of each well at 550 nm is read using an ELISA reader. The values for the mean +/- SD of data from replicated wells are calculated, using the formula % T/C (% viable cells treated/control).

10 OD of treated cells
----- x 100 + % T/C
OD of control cells

The concentration of test compound which gives a T/C of 50% growth inhibition was designated as the IC_{50} value.

B. In vivo methodology

Compounds of this invention were further tested in pre-clinical assay for in vivo activity which is indicative of clinical utility. Such assays were conducted with nude mice into which tumor tissue, preferably of human origin, had been transplanted (xenografted), as is well known in this field. Test compounds were evaluated for their anti-tumor efficacy following administration to the xenograft-bearing mice.

25 More specifically, human breast tumors (MX-1) which had been grown in athymic nude mice were transplanted into new recipient mice, using tumor fragments which were about 50 mg in size. The day of transplantation was designated as day 0. Six to ten days later, mice were treated with the test compounds given as an intravenous injection or orally, in groups of 5-10 mice at

each dose. Compounds were given every other day, for 3 weeks, at doses from 1-200 mg/kg body weight.

Tumor diameters and body weights were measured twice weekly. Tumor volumes were calculated using the diameters measured with Vernier calipers, and the formula

(Length x width 2)/2 = mm 3 of tumor volume

Mean tumor volumes are calculated for each treatment group, and T/C values determined for each group relative to the untreated control tumors.

The new compounds possess good tumor inhibiting properties.